

## Certificate of Analysis

### PCR Master Mix

Cat. #	Size
M7501	10 reactions
M7502	100 reactions
M7505	1,000 reactions

**Description:** PCR Master Mix includes Nuclease-Free Water and PCR Master Mix, 2X. PCR Master Mix is a premixed, ready-to-use solution containing *Taq* DNA polymerase, dNTPs, MgCl<sub>2</sub> and reaction buffers at optimal concentrations for efficient amplification of DNA templates by PCR.

**PCR Master Mix, 2X:** 50 units/ml of *Taq* DNA polymerase supplied in a proprietary reaction buffer (pH 8.5), 400μM dATP, 400μM dGTP, 400μM dCTP, 400μM dTTP, 3mM MgCl<sub>2</sub>.

**Storage Conditions:** See the Product Information Label for storage recommendations. Minimize the number of freeze-thaw cycles by storing in working aliquots. Product may be stored at 4°C for up to three months. Mix well prior to use.

## Quality Control Assays

### Activity Assays

**Functional Assay:** PCR Master Mix is tested for performance in the polymerase chain reaction (PCR) using PCR Master Mix, 1X, to amplify a 360bp region of the  $\alpha$ -1-antitrypsin gene from 100 molecules (0.35ng) of human genomic DNA. The resulting PCR product is visualized on an ethidium bromide-stained agarose gel.

***Taq* DNA Polymerase Activity Assay:** *Taq* DNA polymerase activity is confirmed before the enzyme is added to the PCR Master Mix, 2X. The polymerase activity is assayed in 50mM Tris-HCl (pH 9.0); 50mM NaCl; 5mM MgCl<sub>2</sub>; 200μM each of dATP, dGTP, dCTP, dTTP (a mix of unlabeled and [<sup>3</sup>H] dTTP); 10μg activated calf thymus DNA and 0.1mg/ml BSA in a final volume of 50μl.

### Contaminant Assays

**Nuclease Assays:** No contaminating endonuclease or exonuclease activity detected.



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*That's Our PCR Guarantee!*

Product must be within expiration date and have been stored and used in accordance with product literature. See Promega Product Insert for specific tests performed.

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Use of this product for basic PCR is outside of any valid US or European patents assigned to Hoffman La-Roche or Applera. This product can be used for basic PCR in research, commercial or diagnostic applications without any license or royalty fees.

Signed by:

R. Wheeler, Quality Assurance

### PCR Master Mix



REF M7502 LOT 0000321696  
-30°C to -10°C 2020-04-30  
Dispensed Lot#: 0000313969  
100 reactions

For Laboratory Use

Country of Origin: USA

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PEEL  
HERE



ADM7502 00003216967



## Promega

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Part# 9PIM750

Printed in USA. Revised 10/16.

## 1. Description

PCR Master Mix has been optimized for use in routine PCR reactions for amplifying DNA template in the range of 0.2–2kb.

## 2. Product Components

Product	Size	Cat.#
PCR Master Mix	10 reactions	M7501

Each system contains sufficient reagents to perform ten 50µl reactions. Includes:

- 250µl PCR Master Mix, 2X
- 1.25ml Nuclease-Free Water

Product	Size	Cat.#
PCR Master Mix	100 reactions	M7502

Each system contains sufficient reagents to perform one hundred 50µl reactions. Includes:

- 2 × 1.25ml PCR Master Mix, 2X
- 2 × 1.25ml Nuclease Free Water

Product	Size	Cat.#
PCR Master Mix	1,000 reactions	M7505

Each system contains sufficient reagents to perform one thousand 50µl reactions. Includes:

- 1 × 25ml PCR Master Mix, 2X
- 1 × 25ml Nuclease Free Water

## 3. Protocol

1. Thaw the PCR Master Mix at room temperature. Vortex the Master Mix and then spin it briefly in a microcentrifuge to collect the material in the bottom of the tube.
2. Prepare one of the following reaction mixes on ice:

### For a 25µl reaction volume:

Component	Volume	Final Conc.
PCR Master Mix, 2X	12.5µl	1X
upstream primer, 10µM	0.25–2.5µl	0.1–1.0µM
downstream primer, 10µM	0.25–2.5µl	0.1–1.0µM
DNA template	1–5µl	<250ng
Nuclease-Free Water to	25µl	N.A.

### For a 50µl reaction volume:

Component	Volume	Final Conc.
PCR Master Mix, 2X	25µl	1X
upstream primer, 10µM	0.5–5.0µl	0.1–1.0µM
downstream primer, 10µM	0.5–5.0µl	0.1–1.0µM
DNA template	1–5µl	<250ng
Nuclease-Free Water to	50µl	N.A.

### For a 100µl reaction volume:

Component	Volume	Final Conc.
PCR Master Mix, 2X	50µl	1X
upstream primer, 10µM	1.0–10.0µl	0.1–1.0µM
downstream primer, 10µM	1.0–10.0µl	0.1–1.0µM
DNA template	1–5µl	<250ng
Nuclease-Free Water to	100µl	N.A.

## 4. General Guidelines for Amplification by PCR

The following guidelines apply to target sequences between 200 and 2,000bp and are optimal for typical thermal cyclers.

### A. Denaturation

- Generally, a 2-minute initial denaturation step at 95°C is sufficient.
- Subsequent denaturation steps will be between 30 seconds and 1 minute.

### B. Annealing

- Optimize the annealing conditions by performing the reaction starting approximately 5°C below the calculated melting temperature of the primers and increasing the temperature in increments of 1°C to the annealing temperature.
- The annealing step is typically 30 seconds to 1 minute.

### C. Extension

- The extension reaction is typically performed at the optimal temperature for *Taq* DNA polymerase, which is 72–74°C.
- Allow approximately 1 minute for every 1kb of DNA to be amplified.
- A final extension of 5 minutes at 72–74°C is recommended.

### D. Refrigeration

- If the thermal cycler has a refrigeration or "soak" cycle, the cycling reaction can be programmed to end by holding the tubes at 4°C for several hours.
- This cycle can minimize any polymerase activity that might occur at higher temperatures, although this is not usually a problem.

### E. Cycle Number

- Generally, 25–30 cycles result in optimal amplification of desired products.
- Occasionally, up to 40 cycles may be performed, especially for detection of low-copy targets.

## 5. Composition of Buffers and Solutions

### PCR Master Mix

50units/ml	<i>Taq</i> DNA polymerase [supplied in a proprietary reaction buffer (pH 8.5)]
400µM	each: dATP, dGTP, dCTP, dTTP
3mM	MgCl <sub>2</sub>