

## Certificate of Analysis

### PCR Master Mix

Cat.#	Size
M7501	10 reactions
M7502	100 reactions
M7505	1,000 reactions

**Description:** PCR Master Mix includes Nuclease-Free Water and PCR Master Mix, 2X. PCR Master Mix is a premixed, ready-to-use solution containing *Taq* DNA polymerase, dNTPs,  $MgCl_2$  and reaction buffers at optimal concentrations for efficient amplification of DNA templates by PCR.

**PCR Master Mix, 2X:** 50 units/ml of *Taq* DNA polymerase supplied in a proprietary reaction buffer (pH 8.5), 400 $\mu$ M dATP, 400 $\mu$ M dGTP, 400 $\mu$ M dCTP, 400 $\mu$ M dTTP, 3mM  $MgCl_2$ .

**Storage Conditions:** See the Product Information Label for storage recommendations. Minimize the number of freeze-thaw cycles by storing in working aliquots. Product may be stored at 4°C for up to three months. Mix well prior to use.

### PCR Master Mix

REF M7502 LOT 0000306284  
-30°C -10°C  
100 reactions  
Dispensed Lot#: 0000270843  
For Laboratory Use  
Country of Origin: USA

Promega Corporation  
2800 Woods Hollow Road  
Madison, WI 53711-5399 USA



ADM7502 00003062847

PEEL  
HERE

## Quality Control Assays

### Activity Assays

**Functional Assay:** PCR Master Mix is tested for performance in the polymerase chain reaction (PCR) using PCR Master Mix, 1X, to amplify a 360bp region of the  $\alpha$ -1-antitrypsin gene from 100 molecules (0.35ng) of human genomic DNA. The resulting PCR product is visualized on an ethidium bromide-stained agarose gel.

***Taq* DNA Polymerase Activity Assay:** *Taq* DNA polymerase activity is confirmed before the enzyme is added to the PCR Master Mix, 2X. The polymerase activity is assayed in 50mM Tris-HCl (pH 9.0); 50mM NaCl; 5mM  $MgCl_2$ ; 200 $\mu$ M each of dATP, dGTP, dCTP, dTTP (a mix of unlabeled and [ $^3H$ ] dTTP); 10 $\mu$ g activated calf thymus DNA and 0.1mg/ml BSA in a final volume of 50 $\mu$ l.

### Contaminant Assays

**Nuclease Assays:** No contaminating endonuclease or exonuclease activity detected.



## Promega

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Signed by:

R. Wheeler, Quality Assurance

## 1. Description

PCR Master Mix has been optimized for use in routine PCR reactions for amplifying DNA template in the range of 0.2–2kb.

## 2. Product Components

Product	Size	Cat. #
PCR Master Mix	10 reactions	M7501

Each system contains sufficient reagents to perform ten 50µl reactions. Includes:

- 250µl PCR Master Mix, 2X
- 1.25ml Nuclease-Free Water

Product	Size	Cat. #
PCR Master Mix	100 reactions	M7502

Each system contains sufficient reagents to perform one hundred 50µl reactions. Includes:

- 2 × 1.25ml PCR Master Mix, 2X
- 2 × 1.25ml Nuclease Free Water

Product	Size	Cat. #
PCR Master Mix	1,000 reactions	M7505

Each system contains sufficient reagents to perform one thousand 50µl reactions. Includes:

- 1 × 25ml PCR Master Mix, 2X
- 1 × 25ml Nuclease Free Water

## 3. Protocol

1. Thaw the PCR Master Mix at room temperature. Vortex the Master Mix and then spin it briefly in a microcentrifuge to collect the material in the bottom of the tube.
2. Prepare one of the following reaction mixes on ice:

### For a 25µl reaction volume:

Component	Volume	Final Conc.
PCR Master Mix, 2X	12.5µl	1X
upstream primer, 10µM	0.25–2.5µl	0.1–1.0µM
downstream primer, 10µM	0.25–2.5µl	0.1–1.0µM
DNA template	1–5µl	<250ng
Nuclease-Free Water to	25µl	N.A.

### For a 50µl reaction volume:

Component	Volume	Final Conc.
PCR Master Mix, 2X	25µl	1X
upstream primer, 10µM	0.5–5.0µl	0.1–1.0µM
downstream primer, 10µM	0.5–5.0µl	0.1–1.0µM
DNA template	1–5µl	<250ng
Nuclease-Free Water to	50µl	N.A.

### For a 100µl reaction volume:

Component	Volume	Final Conc.
PCR Master Mix, 2X	50µl	1X
upstream primer, 10µM	1.0–10.0µl	0.1–1.0µM
downstream primer, 10µM	1.0–10.0µl	0.1–1.0µM
DNA template	1–5µl	<250ng
Nuclease-Free Water to	100µl	N.A.

## 4. General Guidelines for Amplification by PCR

The following guidelines apply to target sequences between 200 and 2,000bp and are optimal for typical thermal cyclers.

### A. Denaturation

- Generally, a 2-minute initial denaturation step at 95°C is sufficient.
- Subsequent denaturation steps will be between 30 seconds and 1 minute.

### B. Annealing

- Optimize the annealing conditions by performing the reaction starting approximately 5°C below the calculated melting temperature of the primers and increasing the temperature in increments of 1°C to the annealing temperature.
- The annealing step is typically 30 seconds to 1 minute.

### C. Extension

- The extension reaction is typically performed at the optimal temperature for *Taq* DNA polymerase, which is 72–74°C.
- Allow approximately 1 minute for every 1kb of DNA to be amplified.
- A final extension of 5 minutes at 72–74°C is recommended.

### D. Refrigeration

- If the thermal cycler has a refrigeration or "soak" cycle, the cycling reaction can be programmed to end by holding the tubes at 4°C for several hours.
- This cycle can minimize any polymerase activity that might occur at higher temperatures, although this is not usually a problem.

### E. Cycle Number

- Generally, 25–30 cycles result in optimal amplification of desired products.
- Occasionally, up to 40 cycles may be performed, especially for detection of low-copy targets.

## 5. Composition of Buffers and Solutions

### PCR Master Mix

50units/ml	<i>Taq</i> DNA polymerase [supplied in a proprietary reaction buffer (pH 8.5)]
400µM	each: dATP, dGTP, dCTP, dTTP
3mM	MgCl <sub>2</sub>